### **References and Notes**

- Keterences and Notes
  (1) S. Danishefsky, J. Dynak, E. Hatch, and M. Yamamoto, J. Amer. Chem. Soc., 96, 1256 (1974).
  (2) (a) For a recent review of alkaloids based on these ring systems, see J. E. Saxton in the "Alkaloids," Vol. 2, Specialist Periodical Re-ports of the Chemical Society, London, 1972. (b) For recent synthet-ic methods in the pyrrolizidine series, see N. J. Leonard and T. Sato, J. Org. Chem., 34, 1066 (1969); M. Pizzorno, S. M. Albonico, *ibid.*, 39, 731 (1974). (c) For pharmacological properties of pyrrolizidines, see A. B. Pomerov and C. Baper, Eur. (c) Paramacol. 14, 374 see A. R. Pomeroy and C. Raper, Eur. J. Pharmacol., 14, 374 (1971).
- (3)(a) J. D. Roberts and R. H. Mazur, J. Amer. Chem. Soc., 73, 2509 (1951); (b) W. K. Kirmse and D. Grassman, Chem. Ber., 99, 1746 (1966)
- (a) The structure of this compound is consistent with its ir, nmr, and (4)mass spectra low resolution; (b) C, H, and N combustion analyses within 0.4% of theory were obtained for this substance.
- (a) The yields do not include recovered phthalimide olefins in the range of 15-20%; (b) ct. B. W. Peace and D. S. Wulfman, Synthe-(5)137 (1973).
- (6) I. Murakoshi, Yakagaku Zasshi, 78, 598 (1958).
  (7) O. E. Edwards, J. M. Paton, M. H. Benn, R. E. Mitchell, A. Watanatada, and K. N. Vohra, *Can. J. Chem.*, 49, 1648 (1971).
  (8) L. Tenud, S. Farooq, J. Seibi, and A. Eschenmoser, *Helv. Chim. Acta*, 53, 2050 (1970).
  (9) D. Markeberg, J. O. Datata 2007 204 (2007).
- (9) R. J. Mohrbacher, U. S. Patent 3,297,704 (1967); Chem. Abstr., 67, 100, 020h (1967).

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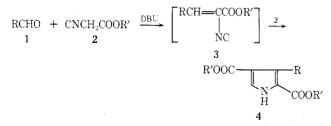
## A Convenient Synthesis of 3-Substituted Pyrrole-2,4-dicarboxylic Acid Esters<sup>1</sup>

Summary: Pyrrole compounds 4 were synthesized by the reaction of isocyanoacetates with aldehydes in the presence of DBU in good yields.

Sir: In the course of our studies on the synthesis of amino acids and related compounds using isocyano compounds, we have investigated the reaction of isocyanoacetates 2 with aldehydes as a source of 3-substituted pyrrole-2,4dicarboxylic acids. Schöllkopf, et al., have detected diethyl 3-methylpyrrole-2,4-dicarboxylate from the reaction of acetaldehyde and ethyl isocyanoacetate in the presence of metallic base during the synthesis of ethyl  $\alpha$ -formylaminoacrylate.2

We have carried out the condensation of alkyl isocyanoacetates with a variety of aliphatic and aromatic aldehydes in THF solution, using 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) as a base (Table I). Spectral and analytical data confirm the pyrrole structures 4. The reaction does not require anhydrous conditions.

The reaction presumably involves Michael addition of isocyanoacetate 2 to the  $\alpha$ -isocyanoacrylate 3; 4 (R = H;  $R' = CH_3$ ) was obtained in 60% yield from the condensation of 2 and methyl  $\alpha$ -isocyanocinnamate.<sup>3</sup> However, the pyrrole was not obtained from the reaction of methyl  $\alpha$ formamidocinnamate with 2.



**Typical Procedure.** To a mixture of methyl isocyanoacetate (1.98 g, 0.02 mol) and DBU (3.0 g, 0.02 mol) dissolved in THF (30 ml) was added dropwise benzaldehyde (1.06 g, 0.01 mol) in THF (10 ml) at 45-50° for a period of 15 min with stirring. After stir-

Table I **Preparation of Pyrrole Derivative 4** 

R	R'	←Condition Temp, °C	ns— Hr	$^{\mathbf{Mp,}}_{^{\circ}\mathbf{C}^{a,b}}$	Yield, %
H	$CH_3$	50-55	1	125-126	67
$CH_3$	$(CH_3)_2CH$	50 - 55	1	74 - 76	71
$CH_{3}CH_{2}$	$CH_3CH_2$	45 - 50	1	87–89°	63
Ph	$CH_3$	45 - 50	5	183 - 185	50
4-Methoxy Ph	$CH_3$	55 - 60	4	146 - 147	57
3,4,5-Tri-					
methoxy Ph	$CH_3$	10 - 15	<b>2</b>	189 - 192	59
3-Pyridine	$\mathrm{CH}_3$	50 - 55	3	212 - 213	60
3-Indole	$CH_3$	50 - 55	3	$>\!250$	58

<sup>a</sup> Recrystallization from aqueous ethanol or methanol. <sup>b</sup> Analyses agreed with the calculated values within  $\pm 0.3\%$ . Lit. mp 88.5–89°: R. Grigg, A. W. Johnson, and T. W. F. Wasley, J. Chem. Soc., 359 (1963).

ring for 5 hr at same temperature, the reaction mixture was neutralized with acetic acid and then the solvent was removed under reduced pressure. The resulting residue was extracted with ethyl acetate and the extract was washed with hydrochloric aicd and water, dried, and then evaporated in vacuo. The crystals (1.3 g) recrystallized from aqueous methanol showed mp 183-185°. The mass spectrum of this compound showed the  $M^+$  at m/e 259 and the ir spectrum (nujol) showed an NH band at 3370 cm<sup>-1</sup> and two ester C=O bands at 1735 and 1700 cm<sup>-1</sup>, respectively. The nmr spectrum (CDCl<sub>3</sub>) indicated the presence of two ester groups (CH<sub>3</sub>) [δ 3.58 (s) and 3.51 (s)], NH [12.04 (br)], pyrrole C-5 H [7.52 (d)], and aromatic H [7.25 (s)].

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### **References and Notes**

- (1) Synthesis of Amino Acids and Related Compounds. 7. Part 6: M. Su-zuki, T. Iwasaki, M. Miyoshi, K. Okumura, and K. Matsumoto, J. Org. Chem., 38, 3571 (1973).
   (2) U. Schölkopf, F. Gerhart, R. Schröder, and D. Hoppe, Justus Liebig Ann. Chem., 766, 116 (1972).
   (3) I. Ugi, U. Fetzer, U. Eholzer, H. Knupfer, and K. Offermann, Angew. Chem., 77, 492 (1965).

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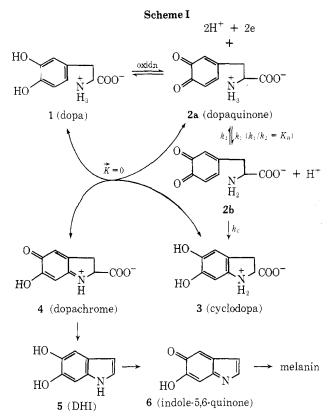
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# Melanin. I. Kinetics of the Oxidative Cyclization of Dopa to Dopachrome

Summary: Kinetics of the ring-closure of dopaquinone (2) to cyclodopa (3) have been studied via chronoamperometry of dopa (1) in the pH range of 5.0-6.0 and at temperatures of 15, 25, 30, and 37°; the rapidity of the cyclization process is attributed to a very favorable positive activation entropy; at pH 6.6 and above, cyclic voltammetry of dopa clearly demonstrates the formation of 5,6-dihydroxyindole (5).

Sir: Biogenesis of the important mammalian pigment melanin<sup>1-4</sup> from 3,4-dihydroxyphenylalanine (1, dopa) has long been considered to proceed via the series of fugitive intermediates illustrated in Scheme I, a pathway originally postulated by Raper<sup>5</sup> and not yet adequately characterized. Mason<sup>6</sup> has estimated the rate constant for decarboxylative rearrangement of dopachrome (4) to 5,6-dihy-



droxyindole (5) to be  $\sim 5.8 \times 10^{-5} \text{ sec}^{-1}$  at pH 5.1. More recently, Brun and Rosset7 have shown by cyclic voltammetry that the redox system,  $1 \Rightarrow 2$ , is essentially reversible in 1 M perchloric acid ( $E_0 = 0.81$  V NHE). Furthermore, above pH 4 dopaquinone (2) cyclizes to cyclodopa (or leucodopachrome, 3), which is then oxidized by dopaquinone (2) to dopachrome (4) yielding a net conversion of 1 to 4 involving four electrons per molecule. This system comprises an ECC mechanism (i.e., an electrochemicalchemical-chemical step sequence) analogous with the oxidative ring closure of adrenalin, whose kinetics have been determined by Adams<sup>8</sup> and coworkers using fast-sweep electroanalytical techniques. We are therefore prompted to report here a similar kinetic study of the dopa  $(1) \rightarrow$ dopachrome (4) conversion at various pH values and several temperatures permitting calculation of the activation-thermodynamic parameters for the cyclization of dopaquinone (2) to cyclodopa (3).

Preliminary cyclic voltammetry experiments were run using a carbon-paste working electrode9 on solutions of dopa (1) at 25°.10 At pH 4.4 and a scan rate of 3.1 V/min dopa showed only a quasi-reversible oxidation-reduction couple with an anodic peak (+0.39 V) for oxidation of 1 and a more cathodic peak (+0.21 V) for reduction of dopaquinone (2). At pH 5.4 incursion of the ring closure of 2 to 3 introduced an additional set of peaks (oxidation, -0.05 V; reduction, -0.21 V) corresponding to the dopachrome (4)-cyclodopa (2) couple. At pH 7.4 the reduction wave for dopaquinone was entirely absent, resulting from rapid cyclization of 2 to 3, the dopachrome-cyclodopa couple was still evident, and a new oxidation peak (+0.05 V) had appeared. For the slow scan rate of 3.1 V/min, this new peak was readily observed at pH 6.6 and above and was in all cases identical with that of an authentic sample of 5,6-dihydroxyindole (5).11

Chronoamperometry experiments following the oxidation of dopa (1) were then run at various pH values and four temperatures (15, 25, 30, and 37°).<sup>10</sup> The current (*i*)-time (*t*) curves were then analyzed using a treatment similar to that of Adams<sup>8</sup> for comparable ECC reactions

Table IFirst-Order Rate Constants for Conversion ofDopaquinone (2a) to Dopachrome (4) at VariousTemperatures<sup>a</sup>

pH		k_ (sec	e <sup>-1</sup> ) at	
	15°	25°	30°	37°
5.00				0.064
5.40	0.014	0.039	0.073	0.13
5.83	0.032	0.099	0.16	0.50
6.00	0.053	0.15	0.24	

<sup>a</sup> The  $k_0$  values represent the average of from three to six individual determinations. Reproducibility varied from 2 to 8% for temperatures of 15 to 30°. Values at 37° are less reliable since the runs at pH 5.83 were recorded on the oscilloscope.

Table IIFirst-Order Rate Constants for Cyclization ofDopaquinone (2b) to Cyclodopa (3) at VariousTemperatures

		k_c (sec <sup>-1</sup> ) at			
$_{\rm pH}$	15°	25°	30°	37°	
5.00				$3.1 \times 10^{2}$	
5.40	$2.7 imes10^{1}$	$7.4 imes10^{1}$	$1.4 imes10^2$	$2.5 imes10^{2}$	
5.83	$2.3 imes10^{1}$	$7.0 imes10^{1}$	$1.1 imes10^2$	$3.6 imes10^{2}$	
6,00	$2.5 imes10^{1}$	$7.2 imes10^{1}$	$1.2 imes10^{2}$		
Av	$2.5 imes10^{1}$	$7.2 imes10^{1}$	$1.2 imes10^2$	$3.1 imes10^2$	
Av %					
Error	5.2	1.8	8.1	12	

of adrenalin and based on the theoretical relationships developed by Hawley and Feldberg.<sup>12</sup> These latter authors have shown that for a reaction system comparable with that in Scheme I, the apparent number  $(n_{\rm app})$  of electrons transferred increases from an initial value of two for the simple redox system (1 = 2), prior to incursion of the ring closure  $(2b \rightarrow 3)$ , to a final value  $(n_{\rm f})$  of four for the overall reaction  $(1 \rightarrow 4)$ . Intermediate values of  $n_{\rm app}$  at times (t) are related to the initial values  $(n_0)$  by eq 1 where C =

$$\frac{n_{\rm app}}{n_0} = \frac{it^{1/2}/C}{(it^{1/2}/C)_{k_0=0}} \tag{1}$$

[dopa], and the value  $(it^{1/2}/C)_{k_c=0}$  is either a value extrapolated to time zero or one-half the value at  $t = \infty$ . Because of uncertainty in the measurements at long times, we chose to determine the intercepts by computer extrapolation of  $it^{1/2}/C$  at various times to t = 0. Minimally triplicate runs provided reproducible intercepts and derived rate data.

Calculated values of  $n_{app}/n_0$  were converted to  $k_0t$ (where  $k_0$  is the observed first-order rate constant for the overall reaction  $1 \rightarrow 4$ ), using the potentiostatic working curve of Hawley and Feldberg<sup>12</sup> for K = 0 (equilibrium constant for the reaction  $1 + 4 \Rightarrow 2a + 3$ ; cf. Scheme I). Plots of  $k_0t$  vs. t were satisfactorily linear and their slopes afforded the observed rate constants ( $k_0$ ) listed in Table I.

Analysis of the kinetics of Scheme I  $(2a \rightarrow 4)$  using a steady-state approximation for intermediate 2b shows that  $k_0 = k_1k_c/(k_2[H^+] + k_c)$  where  $k_c$  is the specific rate constant for ring closure  $(2b \rightarrow 3)$  assuming first-order cyclization. At high  $[H^-]$  where  $k_2[H^+]$  is much greater than  $k_c$  the equation simplifies to eq 2 or 3 where  $K_a$  is the second ionization constant (for  $-NH_3^+$ ) of the dopaquinone

$$k_{0} = \frac{k_{1}k_{c}}{k_{2}[\mathrm{H}^{*}]} = k_{c}\frac{K_{a}}{[\mathrm{H}^{*}]}$$
(2)

$$\log k_0 = pH + \log K_a k_c \tag{3}$$

(2a). As a first approximation this  $K_a$  is taken to be equal to the second ionization constant  $(pK_a = 8.68)$  for dopa (1).<sup>13</sup> Plots of the pH-rate profile at the various temperatures all show approximately the theoretical slope = 1 expected from eq 3 in the pH range of 5.00-6.00. Using data only from this range, the slope of eq 2 yielded the values of  $k_{\rm c}$  summarized in Table II.

An Arrhenius plot (correlation coefficient of 0.996) of the average  $k_c$  values from Table II yields an experimental activation energy of 20.0 kcal/mol for the cyclization of dopaquinone (2b) to cyclodopa (3). The activation thermodynamic parameters calculated at 25° are as follows:  $\Delta H^* = 19.4 \text{ kcal/mol}, \Delta G^* = 14.9 \text{ kcal/mol}, \text{ and } \Delta S^* =$ +15.1 eu. The rapidity of the ring closure may therefore be attributed to a very favorable probability factor as reflected by the large positive entropy of activation. This result is in accord with recent kinetic studies by Illuminati and coworkers,<sup>14</sup> who reported that ring-closure reactions of  $o-\omega$ -bromoalkoxyphenoxides showed a gradual transition from negative to positive entropies of activation as the product ring size decreased from ten to six.

Finally it should be noted that the rate of cyclization of dopaquinone to cyclodopa at 25° is roughly 10<sup>6</sup> times as fast as the reported<sup>6</sup> rate of decarboxylative rearrangement of dopachrome to 5.6-DHI (5). Our cyclic voltammetry experiments suggest that this latter process is much more rapid at physiological pH; hence, additional studies of subsequent steps in Scheme I are currently in progress.

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#### **References and Notes**

- H. S. Mason in "Proceedings of the 4th Congress on Normal and Atypical Pigment Cell Growth," M. Gordon, Ed., Academic Press.
- New York, N. Y., 1959. (2) R. H. Thompson in "Comparative Biochemistry," M. Florkin and H. H. H. Hindpson in Comparing Directionships, Jul. Phys. Rev. Phys. Rev. Press, New York, N. Y., 1962.
   G. A. Swan, Ann. N. Y. Acad. Sci., 100, 1005 (1963).
   H. S. Mason in "Advances in Biology of the Skin," Vol. VIII, Pergamon Press, Oxford, 1967.

- (i) In or Press, Oxford, 1967.
  (5) H. S. Raper, *Biochem. J.*, 20, 735 (1926); 21, 89 (1927). W. L. Duliere and H. S. Raper, *ibid.*, 24, 239 (1930).
  (6) H. S. Mason and C. I. Wright, J. Biol. Chem., 180, 235 (1949).
  (7) A. Brun and R. Rosset, R. Acad. Sci., Ser. C, 275, 1271 (1972); J. Electroanal. Chem., 49, 287 (1974).
  (8) M. D. Hawley, S. V. Tatawadi, S. Piekarski, and R. N. Adams, J. Amer. Chem. Soc., 89, 447 (1967); *ibid.*, 90, 1093 (1968).
  (9) R. N. Adams, "Electrochemistry at Solid Electrodes," Marcel Dekker, Inc., New York, N. Y., 1969, Chapter 9.
  (10) Cyclic voltammetry experiments were performed on racemic dopa using a Wavetek Model 112 triggered VCG and a Wenking Model 68-FR potentiostat. The cell comprised a sce reference electrode, platinum counter electrode, and a carbon-paste working electrode platinum counter electrode, and a carbon-paste working electrode renewed after each run. Output was recorded on a Hewlett-Pack-ard Model 7001A X-Y recorder. Chronoamperometry runs were made with the same equipment using a square wave output and the time-sweep generator of the X-Y recorder. Standard McIlvaine buffers were used. Kinetic runs were made on solutions thermostated to  $\pm 0.04^\circ$ . Data treatment was performed on a CDC-6400 computer. High speed CV and CA runs were recorded photographically on a Tektronix Model 564 storage oscilloscope.
- 5,6-Dihydroxyindole was prepared as described by J. D. Bu'Lock and J. Harley-Mason, *J. Chem. Soc.*, 2248 (1951), and sublimed before use: mp 137-139°; lit. mp 140° dec. M. D. Hawley and S. W. Feldberg, *J. Phys. Chem.*, **70**, 3459 (11)
- (12)(1966)
- (1966). Dopa has  $pK_{a1} = 2.32$ ,  $pK_{a2} = 8.68$ ,  $pK_{a3} = 9.88$ ; *cf.* G. Kortum, W. Vogel, and K. Andrussow, "Dissociation Constants of Organic Acids in Aqueous Solution," Butterworths, London, 1961, p 421. G. Illuminati, L. Mandolini, and B. Masci, J. Amer. Chem. Soc., 96, (13)
- (14)422 (1974)
- (15) Based largely on the doctoral research of J. R. Griswold.

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